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Chemerin as a Biomarker for Periodontitis in Systemically Healthy and Type 2 Diabetic Patients

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Abstract

Aim of the study: The aim of the study was to evaluate the salivary levels of chemerin in patients with periodontitis and type 2 diabetes mellitus.

Materials and methods: A total of 88 subjects were divided into four groups; 13 systemically healthy individuals having a healthy periodontium (control group), 25 patients with type 2 diabetes mellitus and healthy periodontium (T2DM group), 25 patients with generalized periodontitis (P group), 25 patients with generalized periodontitis and type 2 diabetes mellitus (P-T2DM group). The clinical periodontal parameters including plaque index (PLI), bleeding on probing (BOP), probing pocket depth (PPD), and clinical attachment loss (CAL) were determined. Unstimulated saliva was gathered, and the concentration of chemerin was estimated utilizing an enzyme-linked immunosorbent assay.

Results: The results showed that the mean concentrations of chemerin were highest in the P-T2DM group with a mean (of 1.598 ± 0.1499 ng/ml) followed by the P group (1.445 ± 0.1021 ng/ml), T2DM group (1.368 ± 0.096 ng/ml), and control group (1.229 ± 0.152 ng/ml), respectively. There was a statistically significant difference among groups at ($p < 0.05$). For the correlation of chemerin with clinical periodontal parameters, chemerin correlated with PPD in the P group at $p = 0.014$, and chemerin correlated with clinical periodontal parameters PLI, BOP, and PPD in the P-T2DM group at ($p = 0.006$, $p < 0.000$, $p = 0.035$), respectively.

Conclusion: Periodontitis and type 2 diabetes mellitus caused an increase in salivary chemerin concentration, which might be used as a marker for both diabetes and periodontitis.

Keywords: Chemerin, Periodontitis, Type 2 Diabetes Mellitus

INTRODUCTION

Periodontitis is the most widespread health problem that affects esthetic and social life and can lead to tooth loss. It is a multifactorial chronic inflammatory disease that is characterized by increased attachment loss, bone resorption, pocket formation, and gingival bleeding ⁽¹⁾.

Periodontitis is a multifactorial disease, resulting from a mix of numerous variables such as inherited, environmental, and host immunological responses. The biofilm that accumulated on the tooth structure is considered as a primary etiologic factor of periodontitis. The host reaction, on the other hand, influences the disease progression ⁽²⁾.

Diabetes mellitus type II (T2DM) is a global disease characterized by insulin secretion defects and/or a decrease in insulin sensitivity ⁽³⁾. Type 2 diabetes, formerly identified as "noninsulin-dependent diabetes" or "adult-onset diabetes," represents most diabetes cases ⁽⁴⁾.

Diabetes is a risk factor for periodontal disease, and it has been linked to an increase in the incidence, prevalence, and severity of periodontal disease ⁽⁵⁾. Failure of the immune system to remove the cause of inflammation results in a persistent inflammatory response and overexpression of pro-inflammatory cytokines. This chronic inflammatory response is the crucial cause of periodontitis and DM coexistence ⁽⁶⁾.

Chemerin was originally known as tazarotene-induced gene 2 protein or retinoid acid receptor responder 2 ⁽⁷⁾. Chemerin is secreted in an inactive form, as prochemerin, which is activated by different serine proteases that transform prochemerin into the complete active state. Chemerin is released from epithelial cells, fibroblasts, adipose tissue, endothelium, and keratinocytes. Chemerin binds to 3 receptors: chemokine-like receptor 1 (CMKLR1 or chemR23), chemokine CC motif receptor-like 2 (CCRL2), and G protein-coupled receptor 1 (GPCR1). ChemR23 is the sole receptor accountable for chemoattraction activity ⁽⁸⁾. Chemerin binding stimulates chemR23 and causes chemoattraction of immune cells to the infected areas ⁽⁹⁾. In addition to stimulating chemotaxis, chemerin has been shown to facilitate macrophage adherence to tissue endothelium ⁽¹⁰⁾. Chemerin may contribute to periodontal tissue deterioration by increasing the expression of pro-inflammatory cytokines and matrix metalloproteinase. The effect of non-surgical periodontal therapy on chemerin levels was considerably decreased in levels of chemerin at 6 months than the baseline values ⁽¹¹⁾. The present study was conducted to evaluate the salivary levels of chemerin in patients with periodontitis and type 2 diabetes mellitus.

MATERIALS AND METHODS

This study was an observational case-control study that included 88 subjects with an age range between (35 - 65) years old. The subjects recruited for the study were patients attending Al-Kindi Teaching Hospital, and Al-Elwya Specialist Dental Center from January 2022 to April 2022. Each subject sign to a written consent for their acceptance to participate after fitting the inclusion criteria. The consent thoroughly describes the goals of the research. The protocol was approved by the ethical committee of the College of Dentistry/University of Baghdad and followed the guidelines of Helsinki and Tokyo for humans (Reference no. 456 in 19-1-2022). Sample size calculators were used to calculate sample size at a 95% confidence interval and a 5% error margin.

Grouping of the study sample

Control group: Consisted of 13 subjects without any systemic disease and having a healthy periodontium.

Type 2 diabetes mellitus (T2DM) group: Consisted of 25 subjects diagnosed to have T2DM with HbA1c $\geq 7\%$ and healthy periodontium.

Periodontitis (P) group: Consisted of 25 subjects who have generalized periodontitis without any systemic disease.

Periodontitis - Type 2 diabetes (P-T2DM) group: Consisted of 25 subjects who have generalized periodontitis and were diagnosed to have T2DM with HbA1c $\geq 7\%$.

Inclusion criteria

- Body mass index BMI ranges between 18.50 kg/m² - 24.99 kg/m² ⁽¹²⁾.
- Presence of at least 20 teeth for each subject.
- For the control group: Bleeding on probing less than 10% and Probing pocket depth ≤ 3 mm ⁽¹³⁾.
- For the periodontitis group: A patient is considered periodontitis if interdental CAL is detectable at ≥ 2 non-adjacent teeth, or buccal or oral CAL ≥ 3 mm with pocketing > 3 mm is detectable at ≥ 2 teeth. All cases were generalized ($> 30\%$ of

teeth involved) and unstable (PPD $\geq$ 4mm with BOP or PPD $\geq$ 5mm) ⁽¹⁴⁾.

Exclusion criteria: Patients with systemic diseases rather than T2DM, pregnant women, smoker patients with, a history of periodontal therapies during the last 3 months, antibiotics treatment during the last 3 months, patients receiving insulin treatment, and patients with diabetic complications.

Saliva collection: Unstimulated saliva was collected from participants between 9 a.m. – 11 a.m. The participants must not eat or drink one hour before the sample collection. The patients were instructed to swallow saliva and then tilt their heads to allow saliva to drool passively. All samples were centrifuged at 3000 rpm for 20 minutes by a centrifuge machine (Hettich, Germany). Then saliva was stored at - 20 C until analyzed by the Enzyme-Linked Immunosorbent Assay (ELISA)

Clinical periodontal parameter

The clinical periodontal parameters were assessed by using William's periodontal probe

- Assessment of Soft Deposits by using the Plaque Index (PLI), no plaque (0) score, presence of plaque (1) score ⁽¹⁵⁾.
- Assessment of Gingival Bleeding on Probing (BOP), if there is bleeding within 15-30 seconds, the surface was given a score of 1, and a score of 0 for the non-bleeding surface ⁽¹⁶⁾.
- Assessment of Probing Pocket Depth (PPD)
- Assessment of Clinical Attachment Level (CAL)

The calibration was made to achieve examiner accuracy and repeatability by measuring the clinical periodontal parameters (BOP, PPD, and CAL) for five volunteers until reach the level of agreement (>0.75) by using the Interclass Correlation Coefficient test.

The concentration of chemerin was estimated utilizing an enzyme-linked immunosorbent assay. Laboratory steps were done according to the manufacturer's instructions (MyBioSource). Absorbance and

concentration were determined using a spectrophotometry plate reader (HumaReader HS, Germany) at 450 nm.

Statistics

Description of data, analysis, and presentation was achieved utilizing Statistical Package for Social Science (SPSS version 24). Mean, standard deviation, and percentage were used to describe data and the inferential statistics applied were One Way Analysis of Variance (ANOVA), Games-Howell or Tukey HSD post hoc tests, independent sample t-test, and Pearson correlation (r). The level of significance at ($P < 0.05$ = significant and $P > 0.05$ = non-significant).

RESULTS

Using the shapiro-Wilks test for testing the normality of data, these data were normally distributed. The total mean and standard deviation of the age were (52.863 ± 8.028) ranging between (35 to 65) years old. Regarding gender distribution, the participants were composed of 51 males and 37 females. The participants were distributed according to age and gender throughout the groups as shown in (table1).

The results showed that the highest means of PLI and BOP were recorded in the P-T2DM group, (73.731 ± 13.314) and (64.068 ± 9.965), respectively. ANOVA test revealed a statistically significant difference among groups for both PLI and BOP at ($p < 0.05$) (table 2). Intergroup multiple comparisons for PLI by utilizing Tukey HSD post hoc test a statistically significant difference between all pairs of groups except between control and T2DM group were recorded. Regarding BOP, intergroup multiple comparisons using the Games-Howell post hoc test showed a statistically significant difference between all pairs of study and control groups (table 3).

PPD and CAL mean in P-T2DM were ($5.0970.601$) and ($3.9920.978$), respectively, and were considered higher than those in the P group ($4.7920.501$) and ($3.5790.918$), respectively. Independent Sample T-test

showed a non-significant difference between the P groups and the P-T2DM group for both PPD and CAL (table 2).

The result showed that the mean concentrations of salivary chemerin were highest in P-T2DM (1.598 ± 0.1499 ng/ml), followed by the P group (1.445 ± 0.1021 ng/ml), T2DM group (1.368 ± 0.096 ng/ml), and control group (1.229 ± 0.152 ng/ml), respectively. ANOVA showed a statistically significant difference in concentrations of chemerin among groups at ($p < 0.05$) (table 2). Following intergroup multiple comparisons using the Games-Howell post hoc test, a statistically significant difference in concentrations of chemerin were found between all pairs of study and control group (table 3).

Using Person's Correlation Coefficient (r) for the correlation between the salivary levels of chemerin with clinical parameters, there was a significant moderate positive correlation with PPD in the P group at $p = 0.014$. While for the P-T2DM group there was a significant moderate positive correlation with PLI at $p = 0.006$, BOP at $p < 0.000$, and PPD at $p = 0.035$ (table 4).

DISCUSSION

Chemerin levels in the periodontitis group were higher than those of the control group with statistically significant differences between them ($P < 0.05$). This result is in agreement with Özcan et al ⁽¹⁷⁾ Which is the first study that investigated the level of chemerin in the saliva of periodontitis patients and suggested that chemerin levels gives more specificity than other adipokines in distinguishing destructive periodontal disease. Also in agreement with Özcan et al ⁽¹²⁾ who found a significantly different in salivary chemerin between the control and periodontitis group with a p-value ($P < 0.05$).

Chemerin's main function in inflammation is chemotaxis. It retrieves inflammatory cells to the site of inflammation, such as macrophages and polymorphonuclear leukocytes ⁽¹⁸⁾. Another

function is that it stimulates pro-inflammatory cytokines like IL1 β and TNF- α ⁽¹⁹⁾, which may play a crucial role in periodontitis by stimulating bone resorption and increasing prostaglandin E2 (PGE2) and secretion of collagenases ⁽²⁰⁾. Chemerin causes permanent tissue damage by raising MMP levels ⁽²¹⁾.

Salivary chemerin level in the T2DM group was higher than the control group with a statistically significant difference between them. Diabetes is thought to be an inflammatory disease, with high levels of proinflammatory mediators ⁽²²⁾. A previous study by Bobbert et al ⁽²³⁾ suggested that chemerin can be applied to predicting diabetes mellitus.

The current study revealed that the salivary chemerin level in P-T2DM was higher than the P group with a statistically significant difference. This result was in agreement with previous studies that also found a higher chemerin level in both saliva and GCF of the periodontitis diabetes group than in only the periodontitis group ⁽²⁴⁻²⁶⁾. Chemerin may be involved in the etiology of periodontitis and diabetes mellitus and the relationship between T2DM and periodontitis may also alter chemerin production. Because diabetes affects the immunologically active molecule, increasing cytokines in periodontal tissues ⁽²⁷⁾.

Regarding clinical periodontal parameters, there was a significant difference between the P group and the P-T2DM group concerning PLI, and BOP. These results agree with previous studies by Abdul-wahab and Ahmed; Hassan and Salman ^(28,29) who also found a significant difference. The possible explanation is that diabetes is usually correlated with increasing gingival inflammation in response to bacterial plaque, suggesting that periodontal tissue reacts differently to local factors ⁽²⁸⁾.

Regarding PPD and CAL, the result showed a higher mean value in the P-T2DM group than in the P group with non-significant differences between them. This result is in agreement with Serrano et al ⁽³⁰⁾

who found a non-significant difference in PPD, and CAL between diabetes patients with periodontitis and non-diabetic patients with periodontitis. Many explanations were proposed to explain the diabetes-related periodontal disease and increase in PPD such as a change in the activity of the polymorphonuclear cell, altered host defenses ⁽³¹⁾, and alterations in tissue homeostasis. This is linked to an increase in collagen degradation owing to increased synthesis of MMP ⁽³²⁾. Furthermore, Hyperglycemia may cause an alteration in the subgingival microenvironment which favor the growth of the utmost prevalent pathogenic microbial species ⁽³³⁾.

Conclusion

Chemerin could be used as an indicator of both diabetes and periodontitis as found that these conditions caused an increase in salivary chemerin levels. The limitation of this study did not consider the severity of periodontitis. For future research suggested measuring the level of chemerin concerning the severity of periodontitis.

Conflicts of Interest

The authors reported that they have no conflicts of interest

References

1. Salman, Z., Ghudhaib, K., Fadhil, R. Evaluating Osteocalcin and Osteonectin in serum male patients with type 2 Diabetes mellitus and periodontitis. *Eurasian Chemical Communications*, 2022; 4(4): 295-302.
2. Meyle J, Chapple I. Molecular aspects of the pathogenesis of periodontitis. *Periodontology* 2000. 2015 Oct;69(1):7-17.
3. Balaky HM, Kakey IS. The Key Role of Bone Function Markers in Patients with Type (II) Diabetes Mellitus. *Iraqi Journal of Science*. 2022 Jul 31:2861-72.
4. American Diabetes Association. 2. Classification and diagnosis of diabetes: Standards of Medical Care in Diabetes—2020. *Diabetes care*. 2020 Jan 1;43(Supplement_1):S14-31.
5. Chee B, Park B, Bartold PM. Periodontitis and type II diabetes: a two-way relationship. *International Journal of Evidence-based Healthcare*. 2013 Dec;11(4):317-29.
6. Amr E, Mostafa R, Shaker O. Possible role of gingival crevicular fluid levels of Chemerin and Fibroblast growth factor 21 as biomarkers of periodontal disease in diabetic and non-diabetic patients. A diagnostic accuracy study. *Advanced Dental Journal*. 2019 Jul 1;1(2):52-63.
7. Shin WJ, Zabel BA, Pachynski RK. Mechanisms and functions of chemerin in cancer: potential roles in therapeutic intervention. *Frontiers in immunology*. 2018 Nov 30;9:2772.
8. Bondue B, Wittamer V, Parmentier M. Chemerin and its receptors in leukocyte trafficking, inflammation and metabolism. *Cytokine & growth factor reviews*. 2011 Oct 1;22(5-6):331-8.
9. Wittamer V, Franssen JD, Vulcano M, Mirjolet JF, Le Poul E, Migeotte I, Brézillon S, Tyldesley R, Blanpain C, Detheux M, Mantovani A. Specific recruitment of antigen-presenting cells by chemerin, a novel processed ligand from human inflammatory fluids. *The Journal of experimental medicine*. 2003 Oct 6;198(7):977-85.
10. Hart R, Greaves DR. Chemerin contributes to inflammation by promoting macrophage adhesion to VCAM-1 and fibronectin through clustering of VLA-4 and VLA-5. *The Journal of Immunology*. 2010 Sep 15;185(6):3728-39.
11. Özcan E, Işıl Saygun N, Serdar MA, Umut Bengi V, Kantarcı A. Non-surgical periodontal therapy reduces saliva adipokine and matrix metalloproteinase levels in periodontitis. *Journal of*

- periodontology. 2016 Aug;87(8):934-43.
12. Consultation WE. Appropriate body-mass index for Asian populations and its implications for policy and intervention strategies. *Lancet* (London, England). 2004 Jan 10;363(9403):157-63.
 13. Chapple IL, Mealey BL, Van Dyke TE, Bartold PM, Dommisch H, Eickholz P, Geisinger ML, Genco RJ, Glogauer M, Goldstein M, Griffin TJ. Periodontal health and gingival diseases and conditions on an intact and a reduced periodontium: Consensus report of workgroup 1 of the 2017 World Workshop on the Classification of Periodontal and Peri-Implant Diseases and Conditions. *Journal of periodontology*. 2018 Jun;89:S74-84.
 14. Tonetti MS, Greenwell H, Kornman KS. Staging and grading of periodontitis: Framework and proposal of a new classification and case definition. *Journal of periodontology*. 2018 Jun;89:S159-72.
 15. O'LEARY TJ. The plaque control record. *J. Periodontol.* 1972;43:38.
 16. Newman MG, Takei H, Klokkevold PR, Carranza FA. Newman and Carranza's Clinical periodontology E-book. Elsevier Health Sciences; 2018 May 29.
 17. Özcan E, Saygun NI, Serdar MA, Kurt N. Evaluation of the salivary levels of visfatin, chemerin, and progranulin in periodontal inflammation. *Clinical oral investigations*. 2015 May;19(4):921-8.
 18. Yoshimura T, Oppenheim JJ. Chemokine-like receptor 1 (CMKLR1) and chemokine (C-C motif) receptor-like 2 (CCRL2); Two multifunctional receptors with unusual properties. *Experimental cell research*. 2011 Mar 10;317(5):674-84.
 19. Berg V, Sveinbjörnsson B, Bendiksen S, Brox J, Meknas K, Figenschau Y. Human articular chondrocytes express ChemR23 and chemerin; ChemR23 promotes inflammatory signalling upon binding the ligand chemerin. *Arthritis research & therapy*. 2010 Dec;12(6):1-2.
 20. Gomes FI, Aragão MG, Barbosa FC, Bezerra MM, Pinto VD, Chaves HV. Inflammatory cytokines interleukin-1 β and tumour necrosis factor- α -novel biomarkers for the detection of periodontal diseases: a literature review. *Journal of oral & maxillofacial research*. 2016 Apr;7(2).
 21. Xue Y, Jiang L, Cheng Q, Chen H, Yu Y, Lin Y, Yang X, Kong N, Zhu X, Xu X, Wan W. Adipokines in psoriatic arthritis patients: the correlations with osteoclast precursors and bone erosions. *PLoS One*. 2012 Oct 29;7(10):e46740.
 22. Ameen EM, Mohammed HY. Correlation between Tumor Necrosis Factor- α and Anti-tyrosine Phosphatase with Obesity and Diabetes Type 2. *Iraqi Journal of Science*. 2022 Aug 31:3322-31.
 23. Bobbert T, Schwarz F, Fischer-Rosinsky A, Maurer L, Möhlig M, Pfeiffer AF, Mai K, Spranger J. Chemerin and prediction of Diabetes mellitus type 2. *Clinical endocrinology*. 2015 Jun;82(6):838-43.
 24. Himanshu Aeran, Amrinder Singh Tuli, Rebecca Chowdhry, Anissa Atif Mirza, Raman Kumar, Basant Kaur Aulakh. Effect of non-surgical periodontal therapy on salivary resistin and chemerin levels in type 2 diabetes mellitus with chronic periodontitis. *International Journal of Oral Health Dentistry*, 2022,8(2):128-134.
 25. Doğan ŞB, Ballı U, Dede FÖ, Sertoğlu E, Tazegül K. Chemerin as a novel crevicular fluid marker of patients with periodontitis and type 2 diabetes mellitus. *Journal of periodontology*. 2016 Aug;87(8):923-33.
 26. Patnaik K, Pradeep AR, Nagpal K, Karvekar S, Singh P, Raju A. Human chemerin correlation in gingival

- crevicular fluid and tear fluid as markers of inflammation in chronic periodontitis and type-2 diabetes mellitus. *Journal of Investigative and Clinical Dentistry*. 2017 Feb;8(1):e12181.
27. Taiyeb-Ali TB, Raman RP, Vaithilingam RD. Relationship between periodontal disease and diabetes mellitus: an Asian perspective. *Periodontology* 2000. 2011 Jun;56(1):258-68.
 28. Abdul-wahab GA, Ahmed MA. Assessment of Some Salivary Enzymes Levels in Type 2 Diabetic Patients with Chronic Periodontitis: Clinical and Biochemical Study. *Journal of baghdad college of dentistry*. 2015 Mar;325(2218):1-3.
 29. Hassan RS, Salman SA. Evaluation of Salivary Melatonin and Periodontal Parameters in Type II Diabetic Patients with Chronic Periodontitis: A Comparative Study. *Indian Journal of Public Health Research & Development*. 2019 Oct 1;10(10).
 30. Serrano C, Perez C, Rodríguez M. Periodontal conditions in a group of Colombian type 2 diabetic patients with different degrees of metabolic control. *Acta odontológica latinoamericana*. 2012 Apr;25(1):130-7.
 31. Kornman KS, Page RC, Tonetti MS. The host response to the microbial challenge in periodontitis: assembling the players. *Periodontology* 2000. 1997 Jun;14(1):33-53.
 32. Kumar MS, Vamsi G, Sripriya R, Sehgal PK. Expression of matrix metalloproteinases (MMP-8 and-9) in chronic periodontitis patients with and without diabetes mellitus. *Journal of periodontology*. 2006 Nov;77(11):1803-8.
 33. Hintao J, Teanpaisan R, Chongsuvivatwong V, Ratarasan C, Dahlen G. The microbiological profiles of saliva, supragingival and subgingival plaque and dental caries in adults with and without type 2 diabetes mellitus. *Oral microbiology and immunology*. 2007 Jun;22(3):175-81.

Table 1: Distribution of age and gender among the study group

	Control	T2DM	P	P-T2DM	Total	P-value
Age (Mean±SD)	49.000±8.621	55.280±7.797	51.440±7.730	53.880±7.655	52.863±8.028	0.089 ^a
Gender						
Male (N)	7	15	16	13	51	0.832 ^b
Percentage	53.85%	60%	64%	52%	57.95%	
Female (N)	6	10	9	12	37	
Percentage	46.15%	40%	36%	48%	42.05%	

^aOne Way ANOVA, ^bChi-Square**Table 2: Mean and standard deviation of clinical periodontal parameters and biomarker for study groups**

	Control	T2DM	P	P-T2DM	df	P-value
Clinical parameter						
	Mean±SD	Mean±SD	Mean±SD	Mean±SD		
PLI	24.641±11.864	32.374±10.213	64.112±15.046	73.731±13.314	3	0.000 ^a
BOP	4.531±2.506	7.111±1.419	55.922±10.449	64.068±9.965	3	0.000 ^a
PPD			4.792±0.501	5.097±0.601	48	0.058 ^c
CAL			3.579±0.918	3.992±0.978	48	0.130 ^c
Biomarker						
Chemerin ng/ml	1.229±0.152	1.368±0.096	1.445±0.1021	1.598±0.1499	3	0.000 ^a

^aOne Way ANOVA, ^cIndependent Sample T-test

Table 3: Inter groups multiple comparisons of chemerin and clinical periodontal parameters (PLI, BOP) between all pairs of study groups by using post hoc tests

		PLI ^d		BOP ^e		Chemerin ^e	
		P	Sig	P	Sig	P	Sig
Control	T2DM	.300	NS	.016	S	.037	S
	P	.000	S	.000	S	.001	S
	P-T2DM	.000	S	.000	S	.000	S
P	T2DM	.000	S	.000	S	.044	S
	P-T2DM	.047	S	.034	S	.001	S
P-T2DM	T2DM	.000	S	.000	S	.000	S

^d Tukey HSD, ^e Games-Howell, S = significant, NS = non-significant

Table 4: Person's Correlation Coefficient (r) between clinical periodontal parameters and the salivary levels of chemerin for each study group

	PLI		BOP		PPD		CAL	
	r	p	r	p	r	p	r	p
Control	0.214	0.482 NS	0.020	0.948 NS				
T2DM	0.284	0.169 NS	-0.042	0.840 NS				
P	0.194	0.352 NS	0.196	0.348 NS	0.483	0.014 S	0.353	0.084 NS
P-T2DM	0.530	0.006 S	0.650	0.000 S	0.424	0.035 S	0.260	0.210 NS

S = significant, NS = non-significant